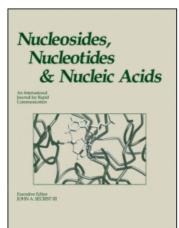
This article was downloaded by:

On: 26 January 2011

Access details: Access Details: Free Access

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



# Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713597286

# 7-Substituted 8-Aza-7-Deazapurines and 2,8-Diaza-7-Deaza-Purines: Synthesis of Nucleosides and Oligonucleotides

Wenqing Linab; Kuiying Xuab; Frank Seelab

<sup>a</sup> Laboratorium für Organische und Bioorganische Chemie, Institut für Chemie, Universität Osnabrück, Osnabrück, Germany <sup>b</sup> Center for Nanotechnology (CeNTech), Münster, Germany

**To cite this Article** Lin, Wenqing , Xu, Kuiying and Seela, Frank(2005) '7-Substituted 8-Aza-7-Deazapurines and 2,8-Diaza-7-Deaza-Purines: Synthesis of Nucleosides and Oligonucleotides', Nucleosides, Nucleotides and Nucleic Acids, 24: 5, 869 — 873

To link to this Article: DOI: 10.1081/NCN-200059218 URL: http://dx.doi.org/10.1081/NCN-200059218

# PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

Nucleosides, Nucleotides, and Nucleic Acids, 24 (5-7):869-873, (2005)

Copyright © Taylor & Francis, Inc. ISSN: 1525-7770 print/ 1532-2335 online

DOI: 10.1081/NCN-200059218



# 7-SUBSTITUTED 8-AZA-7-DEAZAPURINES AND 2,8-DIAZA-7-DEAZA-PURINES: SYNTHESIS OF **NUCLEOSIDES AND OLIGONUCLEOTIDES**

Wenqing Lin, Kuiying Xu, and Frank Seela - Laboratorium für Organische und Bioorganische Chemie, Institut für Chemie, Universität Osnabrück, Osnabrück, Germany and Center for Nanotechnology (CeNTech), Münster, Germany

7-Substituted 8-aza-7-deazaadenosines **1a-e** were synthesized by Sonogashira cross coupling from the corresponding 7-iodo nucleoside in 36-79% yields. Starting from 7-bromo (or 7-iodo)-8-aza-7deazaadenine, 2a,b were obtained by acid-catalyzed glycosylation followed by deprotection in 53 and 35% yields, repectively. Compounds 2b was applied to cross coupling reaction to give 2c-d in 34-95% yield. Compounds 2a and 4b were further transformed to the phosphoramidites 5 and 6b in 9 and 49% overall yields, which were incorporated into oligonucleotides.

Keywords 7-Substituted 8-Aza-7-Deazaadenosine, Phosphoramidite, Sonogashira Cross Coupling, Glycosylation

#### INTRODUCTION

8-Aza-7-deazapurine (pyrazolo[3,4-d]pyrimidine) nucleosides have attracted attention due to their biological activities and as constituents of oligonucleotides<sup>[1,2]</sup> (Scheme 1).

Earlier, 7-substitutents, such as 7-halogeno and 7-alkynyl, have shown favourable effects on the stability of duplex DNA.[3] Herein we report on the synthesis of 7-substituted purine nucleoside analogs 1a-e, 2a-e, 3a,b, and **4a,b**, as well as of the phosphoramidites **5** and **6b**.

The work was financially supported by the European Community (Grant No.: QLRT-2001-00506, "Flavitherapeutics").

Address correspondence to Frank Seela, Laboratorium für Organische und Bioorganische Chemie, Institut für Chemie, Universität Osnabrück, Barbarastr. 7, Osnabrück 49069, Germany.

SCHEME 1

# **RESULTS AND DISCUSSION**

7-Iodo-8-aza-7-deazaadenosine (**7**) was prepared according to Cottam et al., which was applied to Sonogashira coupling reaction with propynes to give **1a,c,d**. Cross-coupling of **7** with trimethylsilylacetylene, followed by desilylation with K<sub>2</sub>CO<sub>3</sub>/MeOH furnished **1b** in 36% overall yield. The ethenyl nucleoside **1e** was obtained by coupling of **7** with tri-n-butyl-vinylstanane under the catalysis of a palladium catalyst and CuI in 74% yield (Scheme 2).

Compound **8a** was prepared from 8-aza-7-deaza-2,6-diaminopurine according to Taylor and Patel. [6] The preparation of **8b** was accomplished by iodination of 8-aza-7-deaza-2,6-diaminopurine with NIS in dichloroethane at refluxing in 65% yield. Glycosylation of **8a,b** with 1-acetyl-2,3,5-tribenzoyl- $\beta$ -D-ribofuranose under catalysis of BF<sub>3</sub>· Et<sub>2</sub>O resulted in **9a,b** in 70 and 53% yield, respectively. Deprotectin of the benzoyl groups in the presence of NaOMe/MeOH gave **2a,b**. Compound **2b** was further applied to cross coupling reaction to furnish **2c-e** in moderate to excellent yield (Scheme 3).

**SCHEME 2** 

#### **SCHEME 3**

The syntheses of  $\mathbf{4a,b}$  were accomplished from  $\mathbf{10a,b}$  via their  $\mathbf{1,N^6}$ -etheno derivatives  $\mathbf{11a,b}$ . Compounds  $\mathbf{10a,b}$  were treated with chloroacetaldehyde in pH  $4{\sim}5$  sodium acetate buffer at r.t. overnight to give  $\mathbf{11a,b}$ . Treatment of  $\mathbf{11a,b}$  with 0.5 M NaOH at ambient temperature overnight yielded  $\mathbf{12a,b}$ , which were cyclized in the presence of NaNO<sub>2</sub> in 80% AcOH to give  $\mathbf{3a,b}$ . Removal of the etheno group of  $\mathbf{3a,b}$  by NBS afforded  $\mathbf{4a,b}$  in 32 and 18% overall yield, respectively (Scheme 4). [7]

Treatment of **2a** with N,N-di-n-butylformamide dimethyl acetal at r.t. gave a mixture of **13/14**, which was further treated with DMTCl in pyridine to yield **15** in 33% overall yield. Because of the unstability of the imino group in N2, the formamide was obtained in these two steps. Treatment of **15** with TBDMSCl and AgNO<sub>3</sub> provided **16** (38% yield), along with its 3'-OTBDMS isomer which could be removed by careful column chromatography. At last, the phosphoramite **5** was obtained in 75% yield (9% overall yield from **2a**) according to the standard procedure (Scheme 5).

Treatment of **4b** with N,N-dimethylformamide dimethyl acetal at r.t. gave **17** in 75% yield. The introduction of the 4,4′-dimethoxytrityl group followed the standard protocol giving compound **18** in 83% yield. Subsequent phosphitylation with chloro-(2-cyanoethoxy)-N,N-(diisopropylamino)phosphine gave the phosphoramidite **6b** in 79% yield (Scheme 6).

Compounds **5** and **6a** were employed in solid-phase oligonucleotide synthesis. Base pairing studies on **4c**, incorporated in a 12-mer duplex, showed that this

### **SCHEME 4**

#### **SCHEME 5**

# **SCHEME** 6

adenine nucleoside analogue forms a strong base pair with dG [5'-d(TAG GTC A4cT ACT)  $\cdot$  3'-d(ATC CAG TGA TGA),  $T_m = 46^{\circ}C$  comparable to 5'-d(TAG GTC AAT ACT)  $\cdot$  3'-d(ATC CAG TGA TGA),  $T_m = 44^{\circ}C$ ] by forming a face to face base pair because of the absence of N7. This novel base pair is as stable as that of the canonical dA-dT pair. However, 4c could not form a stable base pair with dT, dA, and dC.

In conclusion, 8-aza-7-deazapurine nucleosides  ${\bf 1,2}$ , as well as 2,8-diaza-7-deazapurine nucleosides  ${\bf 3a,b}$  and  ${\bf 4a,b}$  were prepared in good to excellent yield. Studies on the biological activities are under way. Compound  ${\bf 2a}$  and  ${\bf 4b}$  were also transformed to the oligonucleotide building blocks  ${\bf 5}$  and  ${\bf 6b}$ . Their incorporation into oligonucleotides is in progress.

## **REFERENCES**

- Seela, F.; He, Y.; He, J.; Becher, G.; Kröschel, R.; Zulauf, M.; Leonard, P. Base-modified oligonucleotides with increased duplex stability: pyrazolo[3,4-d]pyrimidines replacing purines. In *Methods in Molecular Biology*; Herdewijn, P., Ed.; Humana Press Inc.: Totowa, NJ, 2004.
- 2. He, J.; Seela, F. Org. Biomol. Chem  ${\bf 2003},\ {\it 1},\ 1873-1883.$

- 3. Seela, F.; Kröschel, R. Nucleic Acids Res. 2003, 31(24), 7150-7158.
- 4. Cottam, H.B.; Wasson, D.B.; Shih, H.C.; Raychaudhuri, A.; Di Pasquale, G.; Carson, D.A. J. Med. Chem. **1993**, *36*, 3424–3430.
- 5. Sonogashira, K.; Tohda, Y.; Hagihara, N. Tetrahedron Lett.  ${\bf 1975},\,4467-4470.$
- 6. Taylor, E.C.; Patel, H.H. Tetrahedron 1992, 48, 8089-8100.
- 7. Seela, F.; Lindner, M.; Glaçon, V.; Lin, W. J. Org. Chem. 2004, 69, 4695-4700.